Reagents area

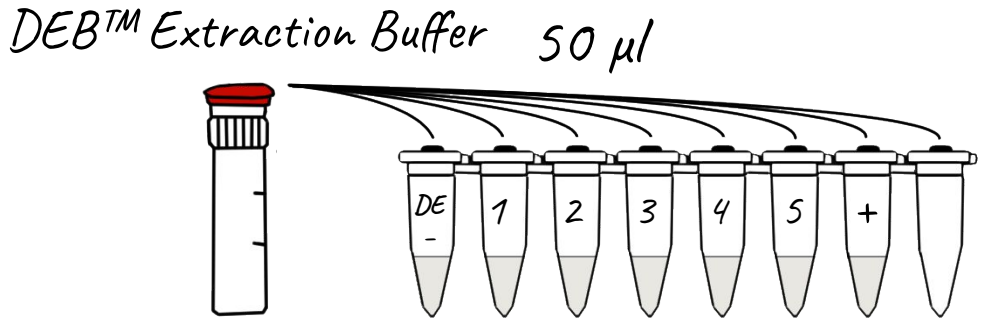
Purpose	• Preparing DEB™ Extraction Buffer • Preparing Complete Master Mix
Precautions	**No samples or completed PCR reactions** should be brought to this area
Materials	*Reagents* DEB™ Extraction Buffer PCR Master Mix Primers *Equipment* 20-200 µl & 100-1000 µl micropipettes Marker miniRack Microtube rack Trash bin Bottle with bleach solution *Consumables* Tissue paper Gloves 200 µl & 1000 µl filter tips PCR tubes 1.5 ml tubes

Reagents area – Materials

Reagents area – Setup

DNA extraction

- Add 50 µl of DEB™ Extraction Buffer to each PCR tube
- One per sample and two for the controls

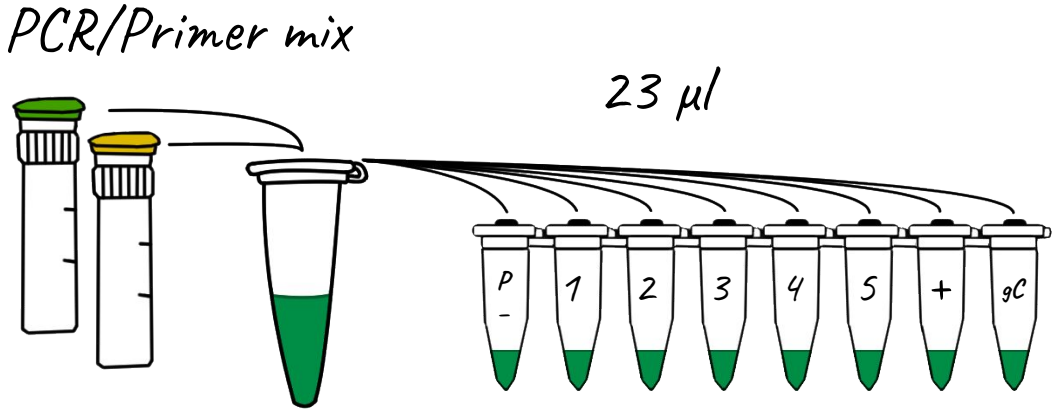


PCR

1. Combine the PCR Master Mix and Primers into a 1.5 ml tube and mix well
2. Add 23 µl of the Complete Master Mix to empty PCR tubes and firmly close the caps

	Number of total tests*		
	1	8	16
PCR Master Mix (µl)	5.5	44	88
Primers (µl)	22	176	352
Total volume (µl)	**27.5**	**220**	**440**

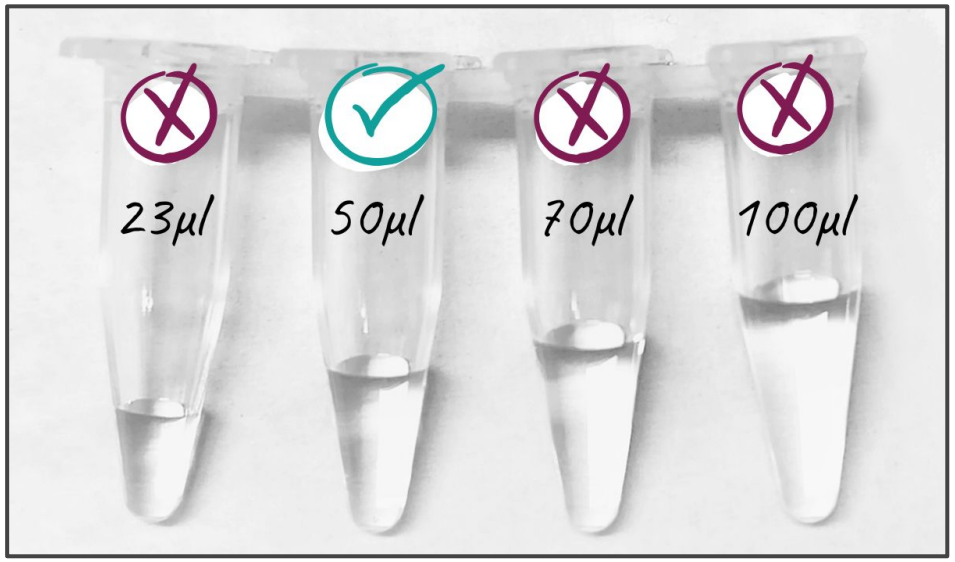
*For other numbers of tests, multiply the volumes required for 1 sample by the total number of tests



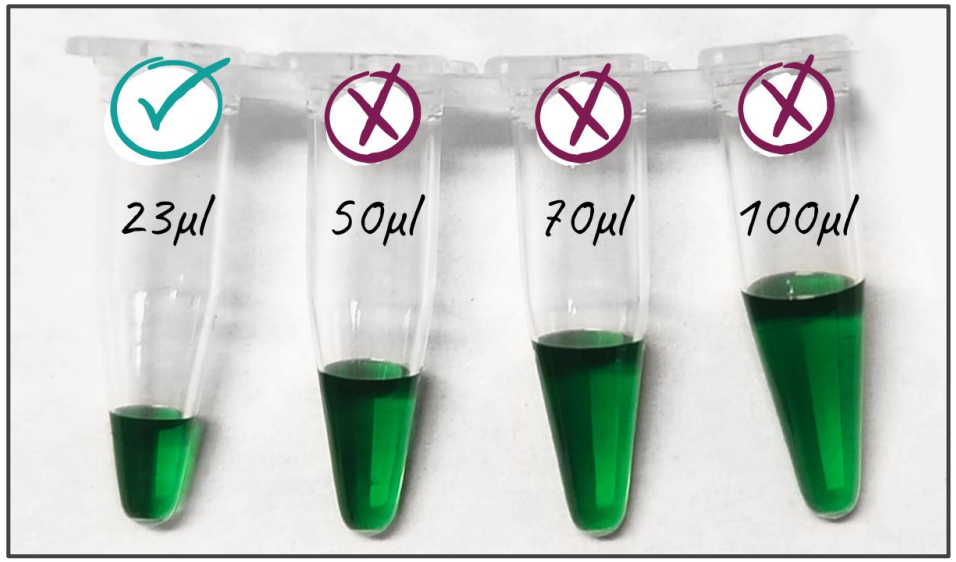
⚠ Clean surfaces with bleach solution!

Reagents area – Volume Reference

DEB™ Extraction Buffer: 50 µl



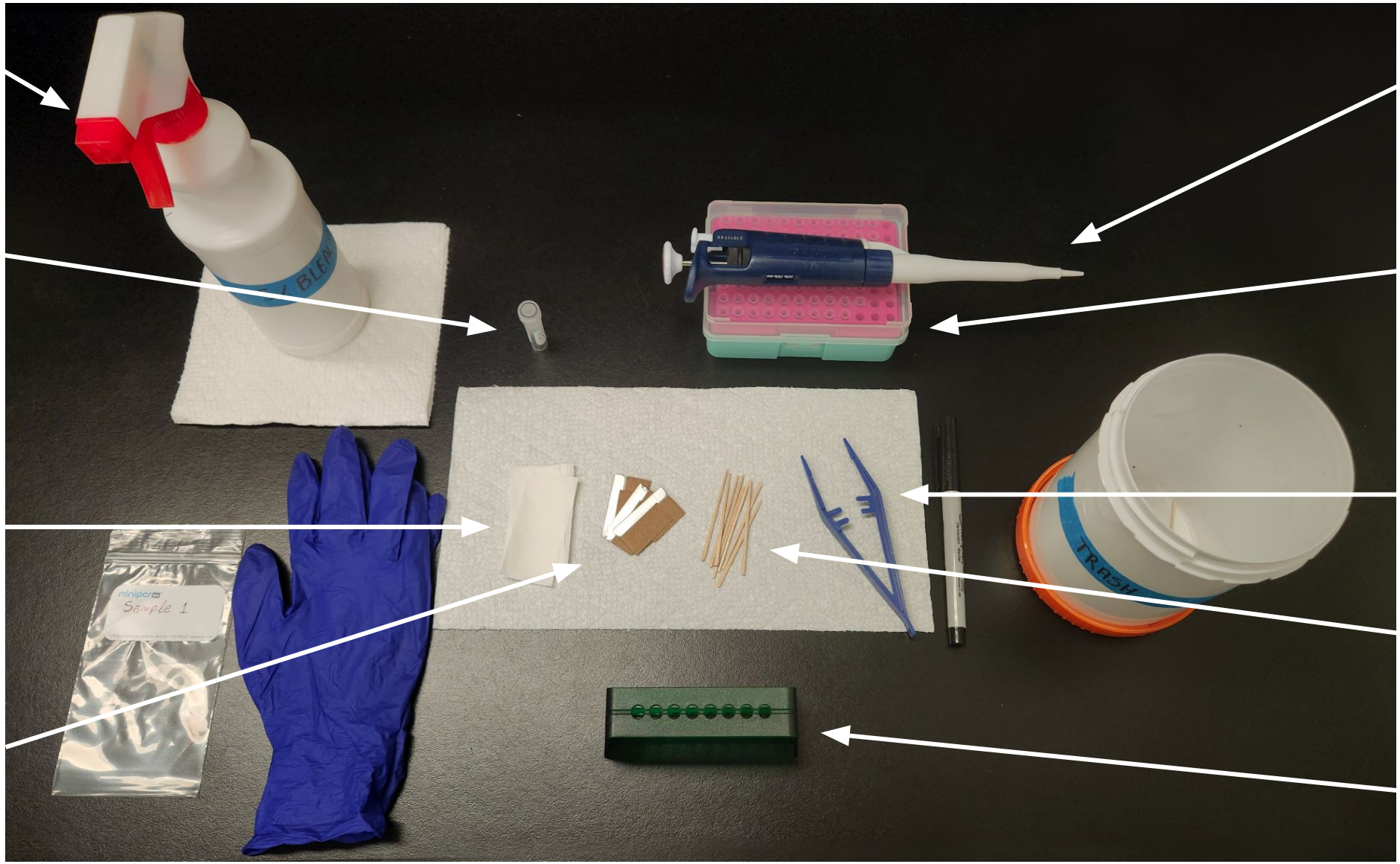
Complete Master Mix: 23 µl



Sampling area

Purpose	• Preparing samples • Transferring lysates to the Complete Master Mix • Transferring control to the Complete Master Mix
Precautions	Material exposed to the Sampling area should **not be returned to the Reagents area**
Materials	*Reagents* gelPCR™ Control 1 *Equipment* Microcentrifuge (*optional*) 1-10 µl micropipette Marker miniRack Trash bin Bottle with bleach solution *Consumables* Tissue paper Gloves 10 µl filter tips Parchment paper Sample collection tools (e.g. punches, razor blades, disposable tweezers, etc.)

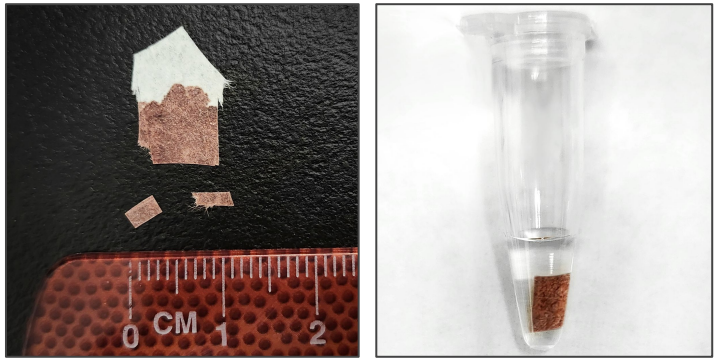
Sampling area – Materials



Sampling area – Lysis

1. Add sample to the DEB™ Extraction Buffer
   (reminder: clean or change gloves in between samples!)
2. Close the cap firmly
3. Dispose of used collection tools
4. Repeat for all samples

Example of sample lysate tube



Move to the PCR area and start the Lysis program

Return to the Sampling area once Lysis is complete

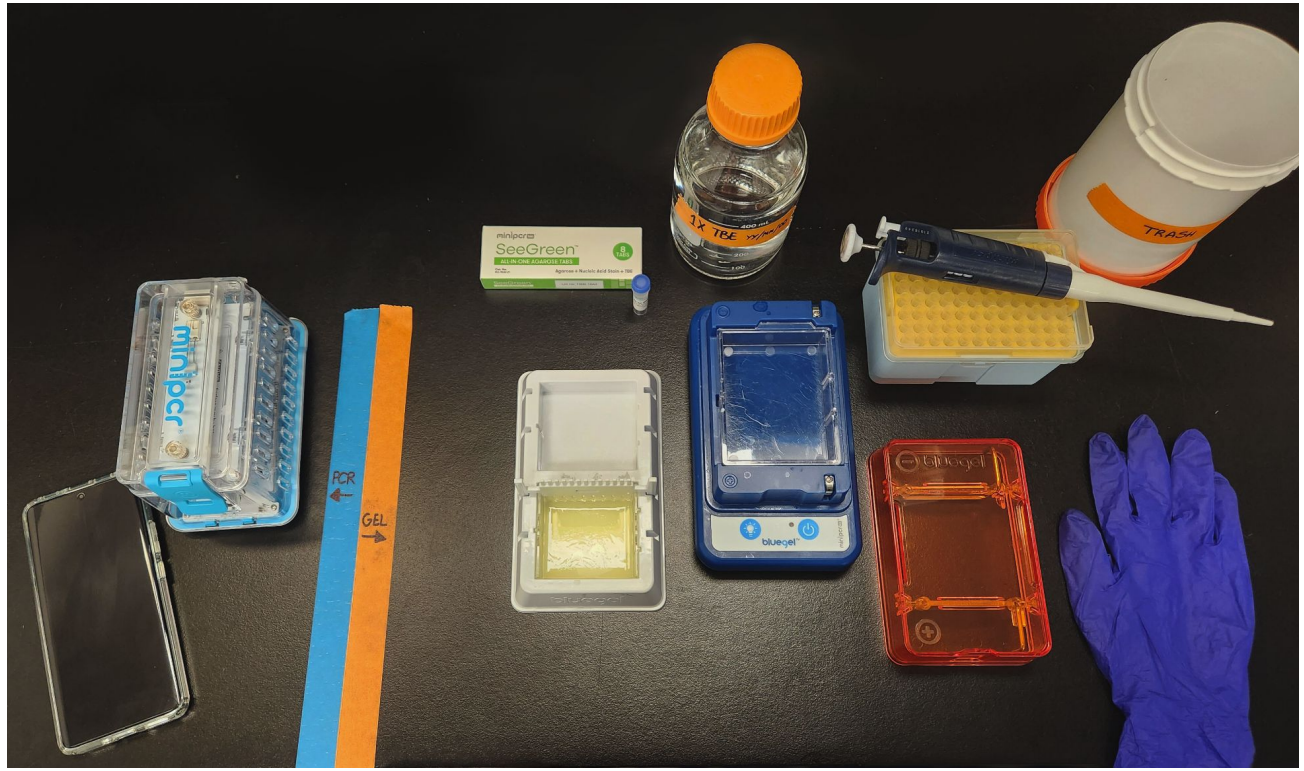
1. Transfer 2 µl of lysate to the corresponding tube with PCR mix
   (reminder: change tip between samples!)
2. Close the cap firmly
3. Repeat for all samples
4. Add 2 µl of gelPCR Control 1 to one tube (optional)

⚠ Clean surfaces with bleach solution!

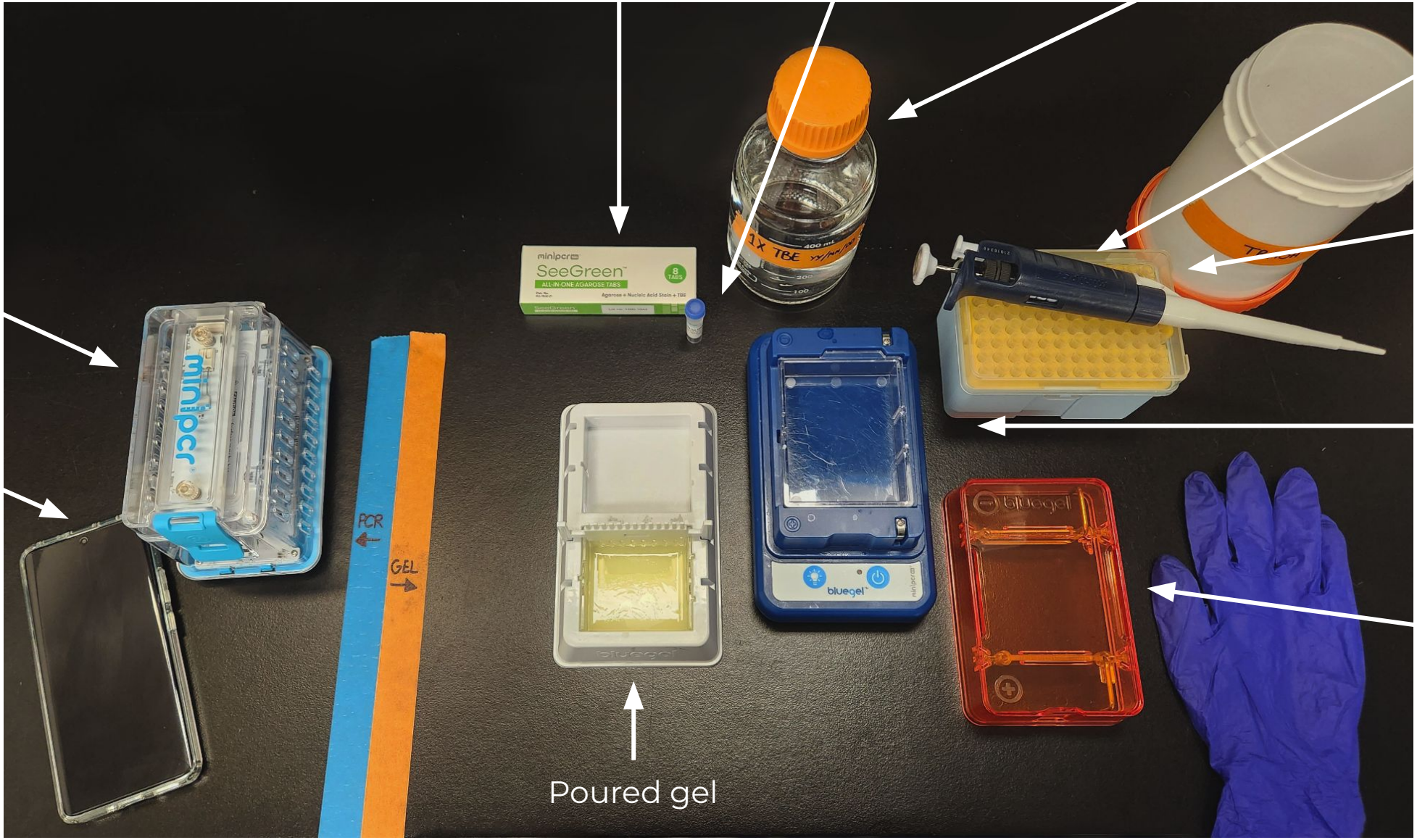
Move to the PCR area and start the PCR program

PCR area

Purpose	• Running PCR • Visualizing test results
Precautions	Material exposed to the PCR area **<u>should not be returned to the Reagents area</u>**
Materials	*<u>Reagents</u>* SeeGreen™ tabs TBE DNA ladder *<u>Equipment</u>* miniPCR® Phone Gel-electrophoresis & visualization system (e.g. blueGel™) 2-20 µl micropipette Trash bin *<u>Consumables</u>* 20 µl filter tips Gloves



PCR area – Materials



PCR area – PCR

1. Load tubes into the miniPCR®
2. Run the specific PCR protocol

How to run miniPCR®

PCR area – Gel Run

1. Prepare gel using SeeGreen™ tabs (reminder: completely dissolve the tab!)
2. Prepare 1X TBE buffer
3. Set up the gel electrophoresis device (if using blueGel™, add 30 ml buffer to the chamber)
4. Load 10 µl of ladder to the first well
5. Load 10 µl of each sample (reminder: change tip between samples!)
6. Run for 15-30 min
7. Take a picture and analyze the results
8. Discard used material

How to make a gel

How to use blueGel™